Evolution and regulation of microbial secondary metabolism

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Pseudomonas aeruginosa

Sensor

sensin

P. aeruginosa

on soft agar

Kinase

Response

Regulator

- Opportunistic pathogen
- Common infections in cancer patients, cystic fibrosis ...
- Biofilm / Swarming behaviour
- Surfactant secretion
- Rhamnolipids



Rhamnolipids

- Secondary metabolites
- (not essential for survival, growth and reproduction)
- Natural surfactants
- Decrease surface tension
 - A. Phenotype assay for P. aeruginosa surfactant secretion



B. Validation on strain PA14 and its engineered mutants



Secondary metabolism

- Relevant for social and ecological interactions
- Surfactant secretion benefits:
 - Motility / Swarming
 - Killing competitors
 - Breaching the epithelial barriers of a host
- Benefit \rightarrow population, Cost \rightarrow individual (rB>C)
- Synthesis is highly regulated (Metabolic Prudence strategy):
 - Quorum sensing
 - Nutrients available (high C/N ratio)

Surfactant production varies inconsistently across the phylogenetic tree

- 28 strains isolated from infected cancer patients (and 3 reference strains)
- Phylogenetic tree from sequence variations in core genome
- Rhamnolipid synthesis genes conserved across all strains



Accessory genome does not explain surfactant secretion phenotype

- Accessory genome gene presence/absence profiles
- 8290 accessory genes, 354 only absent in non-producers
- But no common pattern among non-producers
- No significant differences between surfactant secretion groups



Surfactant secretion lost in strains with slower exponential growth

- Growth on glycerol minimal media
- Growth curves reached similar maximum densities
- Main difference is growth rate in the beginning of exponential phase
- Surfactants are usually produced and secreted only at the end of exponential phase



Metabolomics shows differences associated with loss of surfactant production

- OPLS-DA was able to separate producers from non-producers using metabolite profiles
- 15 metabolites had significant differences between groups
- associated with enzymes containing ROS sensitive Fe-S clusters



Non-producers are worse at reducing oxidative stress

- Strains were exposed to H₂O₂
- H₂O₂ was measured through fluorescent sensor
- Both moderate and high surfactant producers were significantly more efficient at removing H_2O_2



Genome scale metabolic model links oxidative stress, slower growth and lack of surfactants

- Model adapted from P. putida model
- C/N=10 requisite for surfactant production
- Limits growth in normal redox conditions
- Fixed fluxes of NADH, NADPH and GSH synthesis
- High synthesis mimics redox stress
- Rhamnolipid overflow only with redox homeostasis



Explaining Model

- Some strains are not as efficient at maintaining redox homeostasis (why?)
- They require more resources from primary metabolism
- Preventing the activation of rhamnolipid synthesis

A. Rhamnolipid biosynthesis by *P. aeruginosa* requires carbon-rich intermediates from the fatty acid and rhamnose pathways of primary metabolism



B. Lineages that can make rhamnolipids from glycerol exceed all primary needs

C. Lineages that cannot meet primary needs lack rhamnolipid synthesis





Final thoughts

- To fully understand the connection between genotypes and phenotypes we need to understand how they evolved
- Different cellular processes and pathways are highly interdependent
- Some phenotypes can be influenced by genes that we associate to apparently unrelated cellular processes